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RESEARCH ARTICLE



STABILITY INDICATING RP-HPLC METHOD FOR DETERMINATION OF AMISULPRIDE IN TABLET DOSAGE FORM

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A new stability indicating RP-HPLC method was development and validation done for Amisulpride in tablet dosage form. RP-HPLC method was performed for estimation of dosage form and degradants. The method utilizes a mobile phase Acetonitrile:water in the ratio of 60:40 v/v and a flow rate of 1 ml/min with the UV detection at 226 nm for Amisulpride (AMP). The retention time was found to be 4.635 min for Amisulpride. The linearity was found to be in the concentration range of 5-30 µg/ml (r²=0.999) for AMP. Forced degradations were carried out under acid, base, thermal, photolytic and oxidative stress conditions. The method was satisfactorily validated as per the ICH guideline.

Key words: Amisulpride, RP-HPLC, Forced degradation study, Validation, ICH guideline.

INTRODUCTION

Amisulpride (AMS) is chemically, 4-amino-N-{[(2RS)-1-ethyl pyrolidin-2-yl]methyl}-5(ethyl sulphonyl)-2-methoxy benzamide (Figure 1) (Sweetman, 2002; O'neil, 2006; BP, 2006) and used in treatment of schizophrenia. It has high affinity for dopamine D2/D3 receptors. Literature survey revealed different analytical methods for the estimation of Amisulpride in biological systems like HPLC using either UV or fluorescence detection and an IR, UV spectrophotometric and HPLC method are also reported. A UV spectrophotometric method, a chromatographic method and few electrophoretic methods are also reported for the quantitative estimation of Amisulpride in pharmaceutical formulations (Sachse et al 2003; Skibiński et al 2007; Ravisankar and Devala Rao, 2015a; 2015b; Ascalone et al 1996). The stability-indicating assay is a method that is employed for the analysis of stability samples in pharmaceutical industry. Stability testing plays

an important role in the process of drug development. The purpose of stability testing is to provide confirmation on how quality of a drug substance varies with time under the influence of a variety of environmental factors such as temperature, humidity, and light and enables recommendation of storage conditions, and shelf life to be established. The method is expected to allow analysis of individual degradation products (Q1A(R2), 2003; Bakshi and Singh, 2012).







MATERIALS AND METHODS Chemicals

Methanol used was of HPLC grade. All other reagents used were of analytical grade for the forced degradation studies. The pharmaceutical dosage form used in this study was Sulpitac labelled to contain 200 mg of Amisulpride per tablet, purchased from local market.

Apparatus

A RP-HPLC (Shimadzu-20 AD) with PDA detector was utilized. An electronic balance (Roy electronics- LCBCN5) was used for weighing the samples. Hot air oven (Coslab CLE-101) was used for the thermal degradation study. A Sonicator (Labfit) was also used.

Preparation of standard stock solutions

Stock solution of AMP (1000 μ g/ml) was prepared by dissolving 100 mg of AMP in 75 ml of methanol in a 100 ml volumetric flask, volume was made up to the mark with methanol to get solution of strength 1000 μ g/ml. 10 ml of the solution was pipetted out from 1000 μ g/ml solution and transferred to 100 ml volumetric flask and volume was made up with methanol up to the mark, and final dilution was 100 μ g/ml solution of AMP.

Selection of sampling wavelengths

The equivalent of 200 mg of AMP was accurately weighed in 100 ml volumetric flasks separately. After the immediate dissolution, the volume was made up to the mark with solvent. These standard stock solutions were observed to contain 100 μ g/ml of AMP. From the above stock solution, working standard solutions having concentration of 5 μ g/ml was prepared by appropriate dilution. Working standard solutions of 5 μ g/ml of the drug were scanned in the range 400-200 nm in the spectrum mode at the low scan speed to obtain the overlain spectra of these drugs (Figure 2).



Fig. 2. Chromatogram of AMP in mobile phase

Selection of mobile phase and optimization of method

Different column chemistry, solvent type, solvent strength, detection wavelength and flow rate were varied to determine the chromatographic conditions giving the best separation. The mobile phase conditions were optimized so that the components were not interfered with the solvent and excipients. Appropriate k range for eluted peaks, assay sensitivity, solvent noise and use of the same solvent system for extraction of drug from formulation matrices during drug analysis were also considered.

A series of aqueous mobile phases containing methanol, acetonitrile, water were also tested. The best results were obtained when acetonitrile and water solvents were used. Further, the method was optimized by changing the concentration of mobile phase and the results were reported.

From the study, it was found that best result was obtained in a quality separation in terms of peak symmetry, reasonable run time and other parameters by use of 50:50 (v/v) ratio mixture of Acetonitrile:water as mobile phase. The flow rate was determined by testing the effect of different flow rate on the peak area and flow rate of 1 ml/min was found optimum.

Preparation of standard stock solutions for linearity study

From the standard stock solutions of 1000 μ g/ml, different dilutions were prepared for each drug having concentration. Then, 20 μ l of these solutions were injected into the LC system with the help of Hamilton syringe. The chromatograms were recorded at 299 nm. From the chromatogram, it was cleared that AMP retention time was 4.635 min from which their area was noted and calibration curve was plotted between the peak area against their respective concentrations. From the calibration curve, it was cleared that AMP has linearity range between 5-25 μ g/ml (**Figure 3**).

Analysis of tablet formulation

As the result of standard analysis found satisfactory, the method was applied for the quantitative study of this drug in commercially available tablet. For the preparation of the stock solution of tablet dosage form, 20 capsules of Sulpitac were taken and their average weight was determined, after which they were crushed to fine powder. Then, powder equivalent to 40 mg of AMP was taken in 50 ml volumetric flask



Fig. 3. Calibration curve of AMP in mobile phase

and dissolved in 30 ml of methanol with vigorous shaking for 5-10 min. The supernatant liquid was transferred to 100 ml of volumetric flask through a 0.4 μ m membrane filter paper. The residue was washed twice with solvent and the combined filtrate was made up to 100 ml mark. 10 ml of the above solution was diluted up to 100 ml solvent. Six replicate of sample solutions were prepared for required concentrations of the three drugs. Then, 20µl of each replicate were injected into the system and their chromatograms were recorded. From the chromatograms, it was observed that AMP was eluted at 4.635 min. The concentrations of these drugs were extrapolated from their respective calibration curves by using the area.

Recovery study

Linearity and range

The six point calibration curves that were constructed were linear over the concentration range between 5-25 μ g/ml for Amisulpride. This was repeated for 3 times.

Precision

For evaluation of intraday precision, repeatability of the result was evaluated for the concentration of 10 μ g/ml for Amisulpride by 3 replicate determination at interval of 1 hr and for the evaluation of interday precision, repeatability of result was evaluated for concentration of 10 μ g/ml for Amisulpride by 3 replicate determination at interval of 1 hr for 3 days.

Limit of detection

Limit of detection for Amisulpride was found to be $0.31537 \mu g/ml$.

Limit of quantification

Limit of quantification for Amisulpride was found to de $1.05123 \mu g/ml$.

Robustness

Robustness of proposed method was performed by changing the HPLC analyst and remaining condition was keeping constant.

Stability indicating assay method

Acid degradation

In 10 μ g/ml solution, Amisulpride added and kept at room temperature for 24 hr.

Base degradation

In 10 μ g/ml solution, Amisulpride added and kept at room temperature for 24 hr.

Thermal degradation

About 50 mg of drug substance kept at 60°C for 8 hr. Then, the solution was prepared to achieve 10 μ g/ml for Amisulpride.

Photolytic degradation

About 50 mg of drug substance was kept directly to the sun light for 12 hr. Then, the solution was prepared to achieve 10 μ g/ml for Amisulpride.

Statistical analyses

Means, standard deviation (SD), relative standard deviation (RSD) and linear regression analysis were calculated using Microsoft Excel 2007.

RESULTS AND DISCUSSION

Many pharmaceutical compounds undergo degradation during storage or even during the different processes of their manufacture. Several chemical or physical factors can lead to the degradation of drugs. Hydrolysis and oxidation are the most famous chemical degradation routes of drugs.

The main classes of drugs that are subject to degradation are esters, amides and lactams. Ester hydrolysis is frequently base catalyzed, which makes the reaction rapid and irreversible.

In the present work, method development for stability indicating estimation of AMP in tablet dosage form in RP-HPLC was performed by using acetonitrile:water as mobile phase in the ratio of 60:40 (v/v) at a flow rate of 1.0 ml/min and the data are presented in **Table 1**.

In the method, wavelengths utilized for Amisulpride were 226 nm. The percentage recovery value obtained was within standard limit of 98% to 101% for the method which confirmed that the method was accurate and free from any interference of excipients.

Formulation	Drug	Label claim (mg)	% Label claim (Mean± SD)
Tablet	Amisulpride	200 mg	98.79±0.0008

The low value of standard deviation obtained confirmed precision of the method. The reproducibility, repeatability and accuracy of the proposed method were found to be satisfactory. Limit of detection and limit of quantitation were calculated and the result was found to be satisfactory. All recovery studies were compiled in the **Table 2**.

	Mean±SD			
validation parameter	Amisulpride magnesium			
Linearity range	5-25 μg/ml			
Correlation coefficient	0.999			
Slope	32216			
Intercept	23309			
Precision				
Interday				
1 st day	99.8±0.2362			
2 nd day	100.3±0.136			
3 rd day	99.7±0.321			
Intraday				
(1 st hr)	99.8±0.2362			
(2 nd hr)	100.1±0.085			
(3 rd hr)	99.9±0.503			
Recovery				
80%	99.38±0.0036			
100%	100.98±0.0049			
120%	101.96±0.0026			
LOD (mg/ml)	0.31537 μg/ml			
LOQ (mg/ml)	$1.05123 \mu g/ml$			
Robustness	107±0.000974			

Table 2. Validation parameters for RP-HPLC methods

Acidic degradation, alkali degradation, thermal degradation and photolytic degradation were

performed successfully by following ICH guideline Q1A (R2) (**Table 3**).

 Table 3. Forced degradation studies

Condition	Amount found % (μg/ml)	Result (% degradation)
Condition	AMP	AMP
Acid degradation	89.61	10.39
Alkaline degradation	92.35	7.65
Thermal degradation	88.35	11.65
Photolytic degradation	66.36	33.64



Fig. 4. Acid degradation chromatogram of AMP

Degradation study was conducted for Amisulpride in acidic medium and the results are shown in **Figure 4**.

Amisulpride when hydrolyzed with alkali, produced their degradation product and the results are shown in **Figure 5**. **Figure 6** shows the thermal degradation of drug AMP and **Figure 7** shows the photolytic degradation of drug AMP. Multiple peaks were seen at different time interval in all the chromatograms of AMP.



Fig. 5. Alkaline degradation chromatogram of Amisulpride



Fig. 6. Thermal degradation chromatogram of Amisulpride

REFERENCES

- Ascalone V, Ripamonti M, Malavasi B. Stereospecific determination of amisulpride, a new benzamide derivative, in human plasma and urine by automated solid-phase extraction and liquid chromatography on a chiral column. application to pharmacokinetics. *J. Chromatogr. B. Biomed. Appl.* 1996;676(1):95-105. [DOI: 10.1016/0378-4347(95)00418-1]
- Bakshi M, Singh S Development of validated stability indicating assay method- critical review. J. Pharm. Biomed. Anal. 2002;28(6):1011-1040. [DOI: 10.1016/S07 31-7085(02)00047-X]
- British Pharmacopoeia, The Stationary Office, London, 2006;I:149.
- O'neil MJ. The Merck Index An Encyclopedia for Chemicals, Drugs and Biologicals, Merck and Co., Whitehouse Station, NJ. 2006;14:485.
- Q1A (R2). Stability testing of new drug substances and products. International Conference on Hormonization. CPMP/ICH/2736/99, Geneva; 2003.
- Sachse J, Härtter S, Weigmann H, Hiemke C. Automated



Fig. 7. Photolytic degradation chromatogram of Amisulpride

CONCLUSION

All these factors lead to the conclusion that the developed stability indicating RP-HPLC method development is accurate, precise, simple, sensitive and rapid and can be applied successfully for the estimation of Amisulpride in tablet dosage form without interference. The relative standard deviation (RSD) for all parameters was found to be less than one, which indicate that the validity of method are also within the limit so the proposed method can be used for routine quantitative estimation of drug.

determination of amisulpride by liquid chromatography with column switching and spectrophotometric detection. *J. Chromatogr. B. Analyt. Technol. Biomed. Life Sci.* 2003;784(2):405-10. [DOI: 10.1016/s1570-0232(02)008 08-5]

- Skibiński R, Komsta Ł, Hopkała H, Suchodolska I. Comparative validation of amisulpride determination in pharmaceuticals by several chromatographic, electrophoretic and spectrophotometric methods. *Anal. Chim. Acta* 2007;590(2):195-202. [DOI: 10.1016/j.aca.20 07.03.038]
- Sweetman S. Martindale The Complete Drug Reference, Pharmaceutical Press, London. 2002;33:655.
- Ravisankar P, Devala Rao G. Novel RP-HPLC method for the determination of Amisulpride in pure and pharmaceutical formulations. *J. Chem. Pharm. Res.* 2015a;7(6):815-24.
- Ravisankar P, Devala Rao G. Novel spectrophotometric methods for the determination of amisulpride in pure and pharmaceutical formulations. *Int. J. Recent Sci. Res.* 2015b; 6(6):4830-7.
